PATHOLOGY DIAGNOSIS OF CANCER

DIAN YULIARTHA LESTARI

HISTOLOGIC METHODS

PARAFFIN EMBEDDING TECHNIQUE

- \circ Hasil operasi ightarrow diagnosis final
- Biopsi \rightarrow diagnosis pre-operatif; diagnosis final
- Tahapan :
 - Makroskopik ightarrow cut section
 - Mikroskopik : Prosesing \rightarrow embedding \rightarrow potong dan pewarnaan H&E

FROZEN SECTION

- Penderita masih berada dalam ruang operasi (durante operasi)
- Dilakukan pengambilan sedikit jaringan (biopsi)
- Tanpa Fixasi
- Hasil dilaporkan dalam 5-15 menit
 - $\circ \rightarrow$ Representatif / tidak
 - $\circ \rightarrow$ JINAK / GANAS

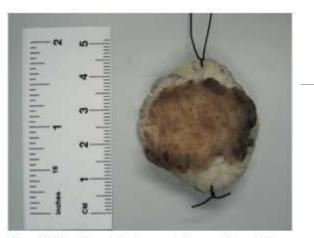


Figure 5.4 A medium-sized skin sample is seen with a central lesion. This could be described as 'A skin ellipse x by y by z mm depth is seen with an orientation suture, designated 12 o'clock. The sample shows a central yellow-brown nodule z mm that is k mm clear of the closest margin'. It is sectioned into parallel slices and then placed into a cassette (see Fig. 5.3).



Figure 5.3 Tissue blocks are placed into the cassette. Note they should not fill the cassette, and must permit room for processing fluid circulation. The orientation of the blocks is enhanced by a sponge securing the specimens in sequential position and a colored agar marker allows designation of the order of slices taken. The samples have been marked with different colored inks to permit designation of the sidedness of the samples and the resection margins.

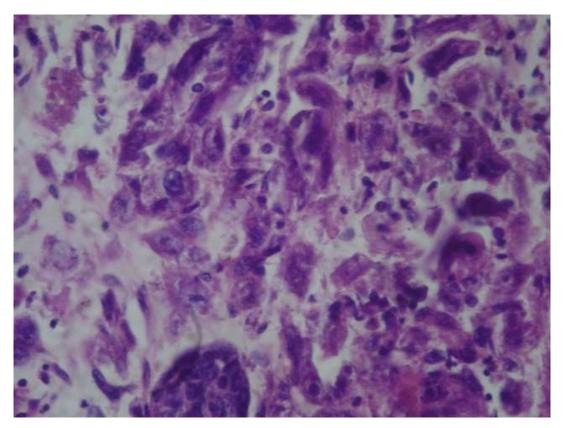


Figure 6.3 Shows an H&E slide and the paraffin block area are matched.

Station	Reagents	Time	P/V	Temp
1	10% Formalin	1 h	Ön	38°C
2	10% Formalin	1 h	On	38°C
2 3 4	50% Alcohol/formalin	1 h	On	38°C
4	70% Alcohol	1 h	On	38°C
5	95% Alcohol	1 h	On	38°C
6	95% Alcohol	40 min	On	38°C
7	100% Alcohal	1.6	On	38°C
8	100% Alcohol	40 min	On	38°C
8 9	Xylene	1 h	On	38°C
10	Xylene	30 min	Ön	38°C
11	Paraffin	30 min	On	60°C
12	Paraffin	30 min	On	60°C
13	Paraffin	30 min	On	60°C
14	Paraffin	30 min	On	60°C

PEWARNAAN

1. Hematoxylin & Eosin



Hematoxylin and eosin stain for paraffin sections

Method

- Dewax sections, rehydrate through descending grades of alcohol to water.
- 2. Remove fixation pigments if necessary.
- Stain in an alum hematoxylin of choice for a suitable time.
- Wash well in running tap water until sections 'blue' for 5 minutes or less.
- Differentiate in 1% acid alcohol (1% HCl in 70% alcohol) for 5–10 seconds
- Wash well in tap water until sections are again 'blue' (10–15 minutes), or
- Blue by dipping in an alkaline solution (e.g. ammonia water), followed by a 5-minute tap water wash.
- 8. Stain in 1% eosin Y for 10 minutes.
- 9. Wash in running tap water for 1-5 minutes.
- 10. Dehydrate through alcohols, clear, and mount.

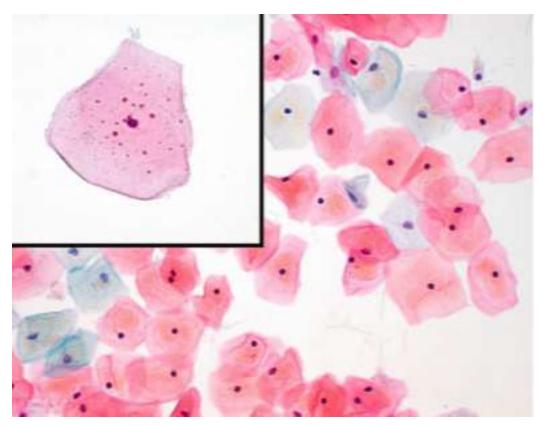
Results

Nuclei Cytoplasm Muscle fibers Red blood cells Fibrin blue/black varying shades of pink deep pink/red orange/red deep pink

Notes

Note that structures and substances other than nuclei may be hematoxyphilic to varying degrees. Examples include fungal hyphae, which are faintly hematoxyphilic, and calcium deposits, which are often deep blue-black.

2. PAPANICULAOU



Papanicolaou staining method

- Remove polyethylene glycol fixative in 50% alcohol, 2 minutes.
- Hydrate in 95% alcohol, 2 minutes, and 70% alcohol, 2 minutes.
- 3. Rinse in water, 1 minute.
- 4. Stain in Harris's hematoxylin, 5 minutes.
- 5. Rinse in water, 2 minutes.
- Differentiate in 0.5% aqueous hydrochloric acid, 10 seconds approx.
- 7. Rinse in water, 2 minutes.
- 8. 'Blue' in Scott's tap water substitute, 2 minutes.
- 9, Rinse in water, 2 minutes.
- 10. Dehydrate, 70% alcohol for 2 minutes.
- 11. Dehydrate, 95% alcohol, 2 minutes.
- 12. Dehydrate, 95% alcohol, 2 minutes.
- 13. Stain in OG 6, 2 minutes.
- 14. Rinse in 95% alcohol, 2 minutes.
- 15. Rinse in 95% alcohol, 2 minutes.
- 16. Stain in EA 50, 3 minutes.

Change stains frequently.

17. Rinse in 95% alcohol, 1 minute.

The staining times can be adjusted to suit personal preference for a darker or paler stain. Alternatives to Scott's tap water substitute include 0.1% ammoniated water or a weak aqueous solution of lithium carbonate.

Results

The nuclei should appear	blue/black
Cytoplasm (non-keratinizing squamous	blue/green
cells)	
Keratinizing cells	pink/orange
Note	

CYTOLOGICAL METHODS

SITOLOGI \rightarrow ilmu yang mempelajari morfologi sel

- $\,\circ\,$ Sitoplasma $\,
 ightarrow\,$ kualitas dan kuantitas
- Nucleus \rightarrow kromatin, ukuran, nucleoli

Pewarnaan yang umum digunakan

- Papaniculaou
- HE

EXFOLIATIVE CYTOLOGY

- Pap smear
- Sitologi urine, pleura, ascites, brochus (washing/brushing)

FINE NEEDLE ASPIRATION CYTOLOGY (FNAC)

Sampel untuk Sitologi

Cervical / Vaginal smear

Sputum

Bronchial washing / brushing

Nasopharyngeal smear/washing/brushing

Urine

Cairan lambung/pleura/ascites/sendi

Liquor serebrospinal

Aspirasi Jarum Halus

Inprint neoplasma

Pembuatan Sediaan

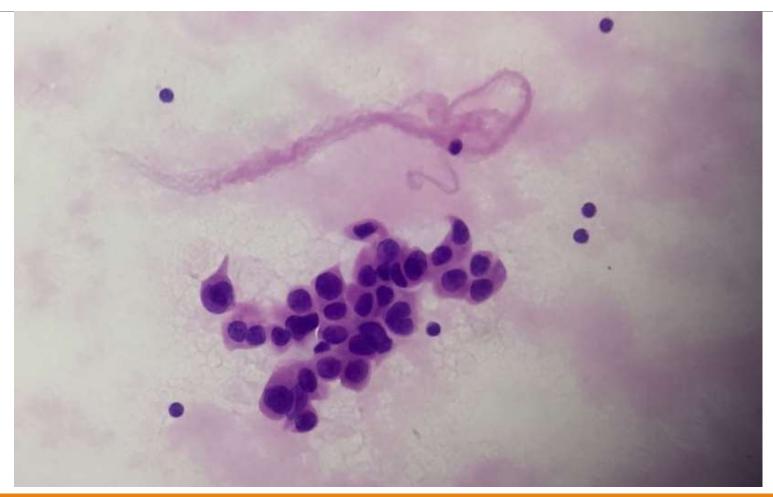
Bahan cair :

Sampel + Fiksatif \rightarrow sentrifuge \rightarrow endapan di smear pada objek glass \rightarrow pengecatan

Cairan : Ascites, Pleura, Urine tamping, BALL, Washing

Pemeriksaan sel-sel yang terlepas

SITOLOGI EKSFOLIATIF



PAP SMEAR

- Pap Smear adalah pemeriksaan sitologi abrasive ginekologi dari serviks untuk deteksi dini kanker serviks
- •1947, seorang ahli gynecology dari Canada J. Ernest Ayre Memperkenalkan alat spatula untuk memperoleh sample Pap Smear lebih baik dan dapat dengan mudah mencapai target. Spatula tersebut dari kayu yang dipotong sesuai bentuk yang diinginkan. Spatula tersebut dikenal sebagai Spatula Ayre.



Tujuan Pemeriksaan Pap Smear

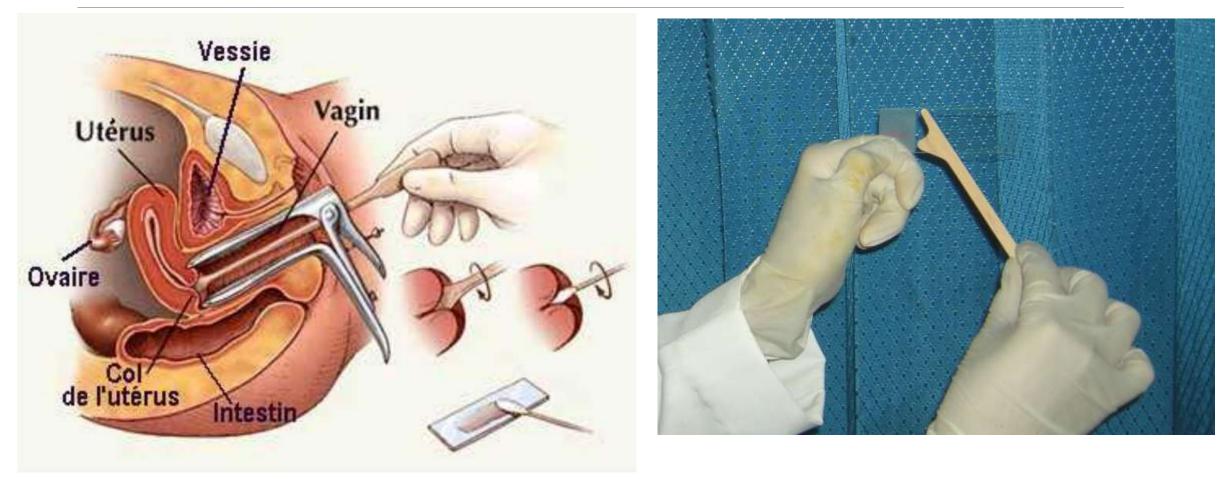
•Deteksi dini kanker serviks \rightarrow proses radang pada serviks dan vagina

Jenís-jenís Pap Smear

Pap smear Konvensional

Pap smear Liquid Base

PAP SMEAR KONVENSIONAL

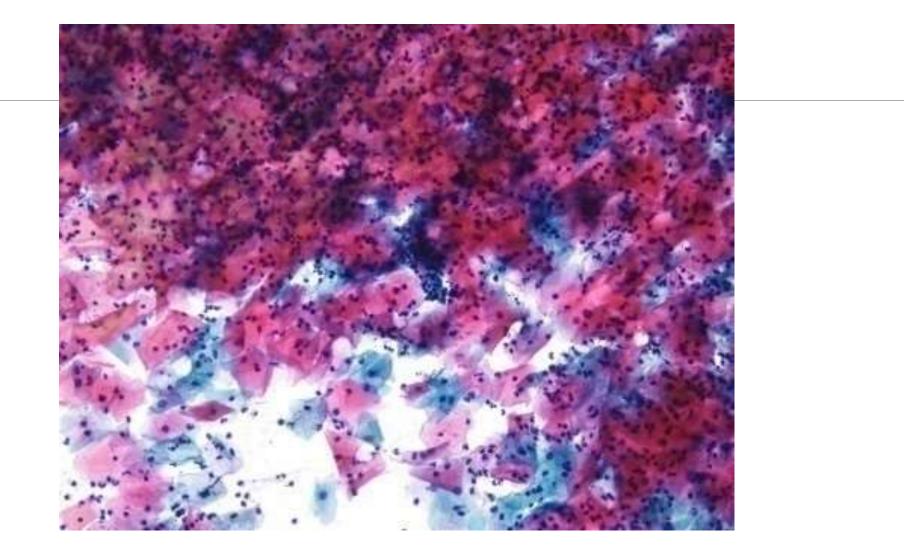


Sel-sel yang diperoleh sering saling menumpuk

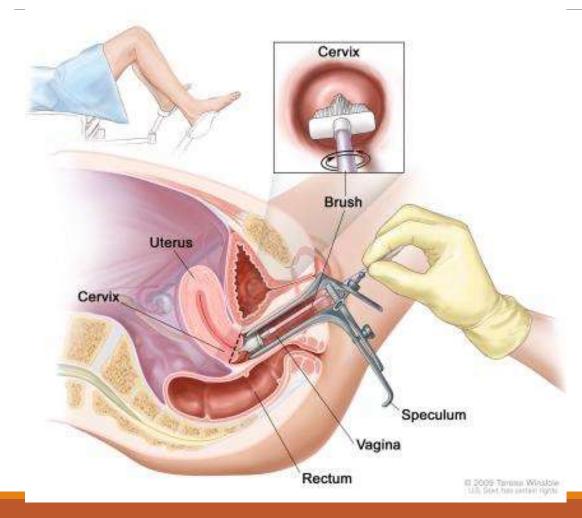
Kabur oleh karena mengandung darah atau lendir.

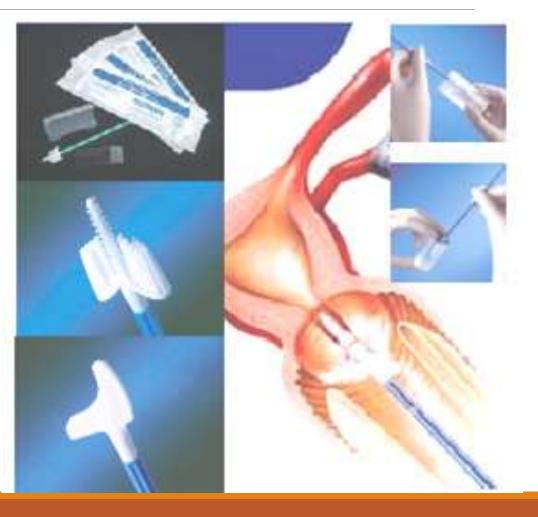
 Selain itu tidak seluruh sample yang diperoleh dilakukan pemeriksaan. Sisa sel yang masih melekat pada spatula akan terbuang.

Akurasi sekitar 76,4 %.



PAP SMEAR LIQUID BASED





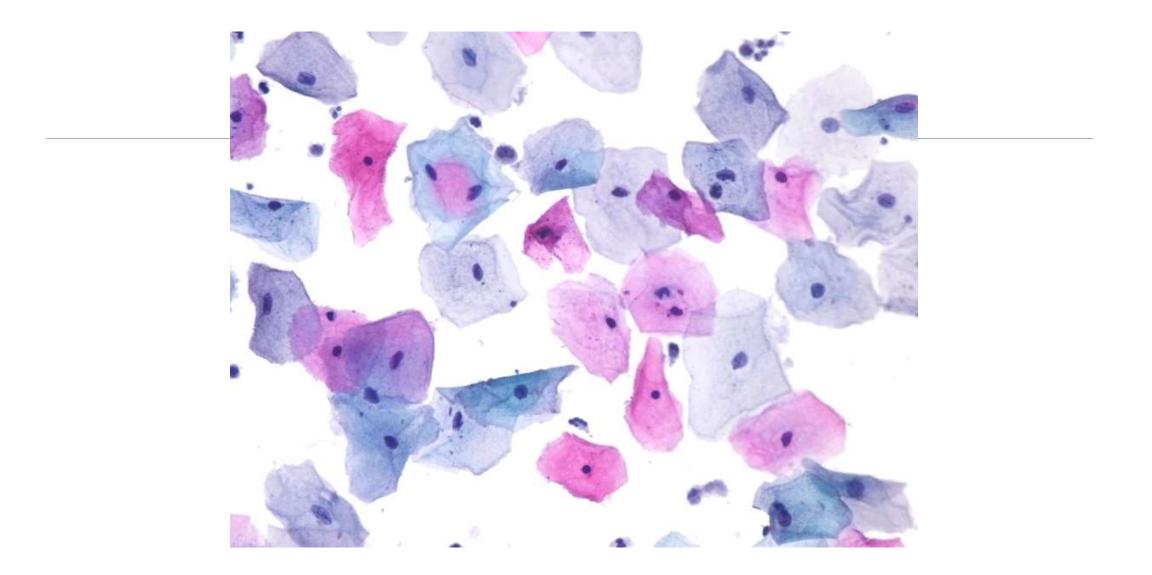
Sel tidak bertumpuk

Latar belakang lebih bersih

Seluruh sel yang diperoleh akan diproses dan tidak terbuang.

Evaluasi tambahan lebih lanjut.

Akurasi sekitar 98 %.



FNAB

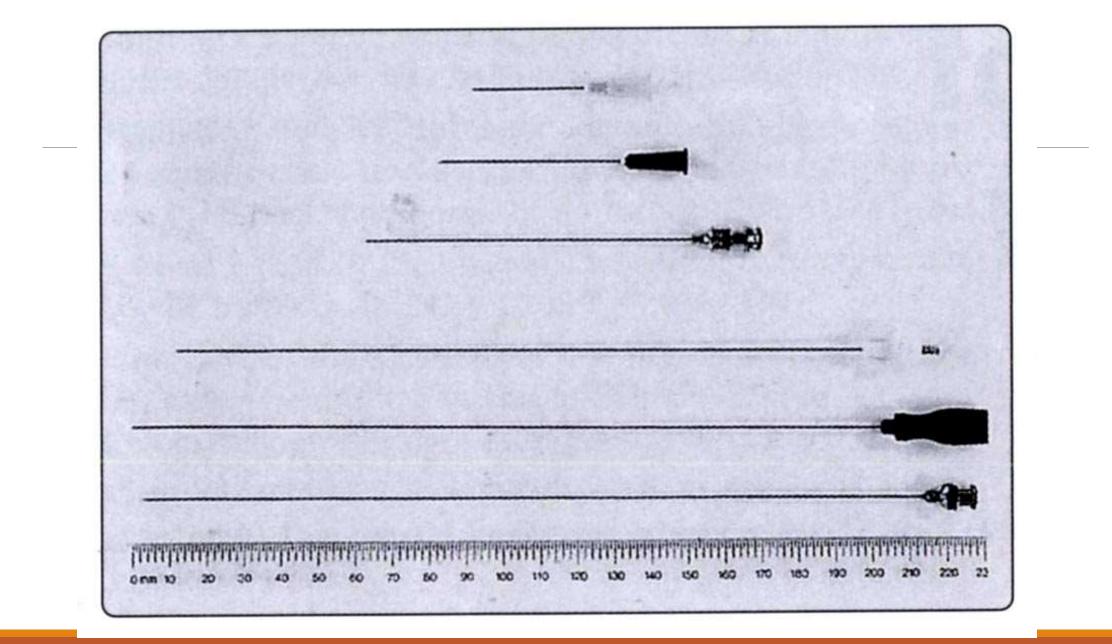
Jarum Halus \rightarrow 25 G atau lebih besar (26G, 27G)

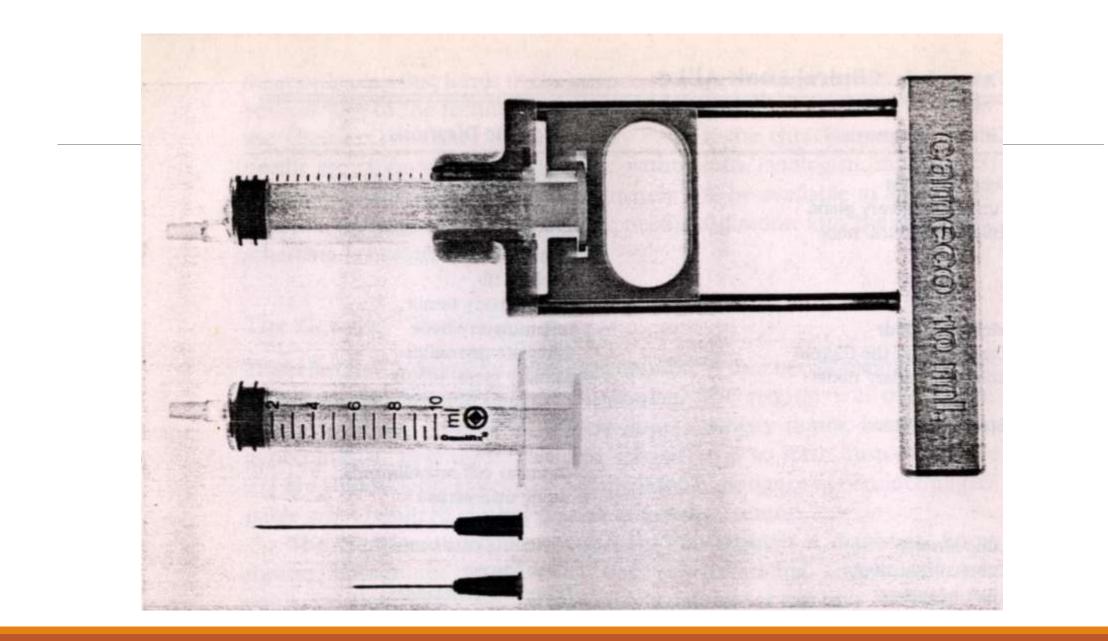
Dapat Dilakukan pada :

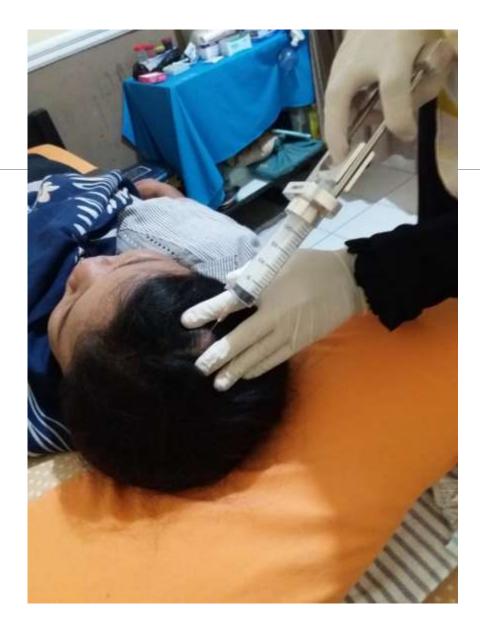
- Tumor-tumor permukaan
- Tumor organ dalam
 - \rightarrow dengan tuntunan CT scan, USG

Keuntungan :

- Tidak traumatik
- Tidak perlu anestesi
- Tidak perlu ruangan khusus
- \rightarrow Diagnosis Pre Operatif

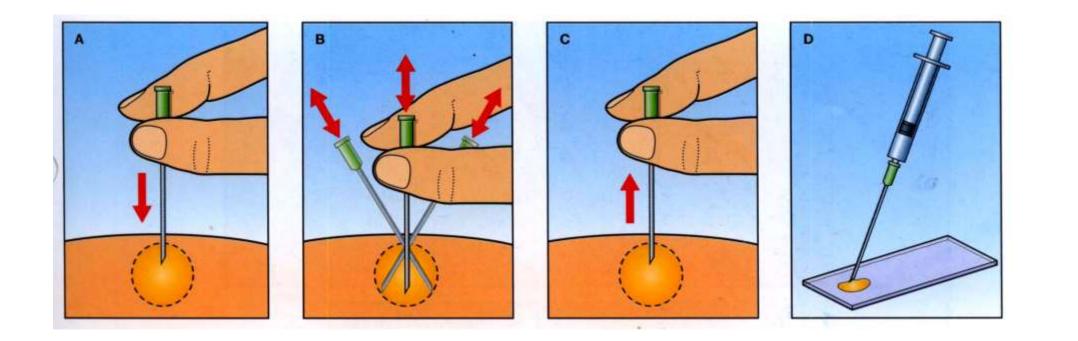


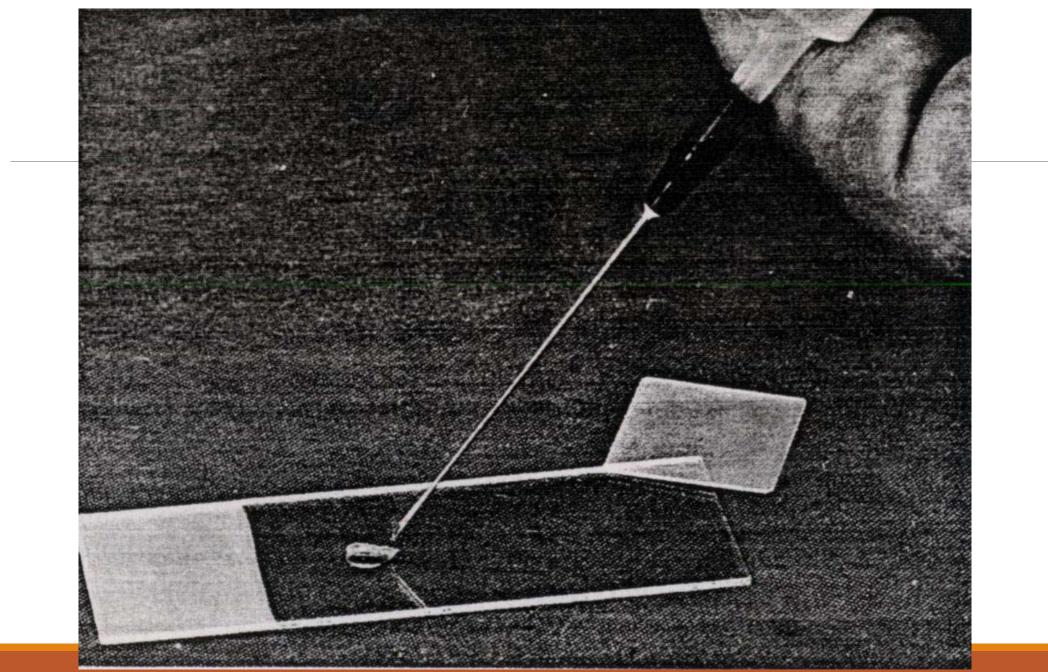


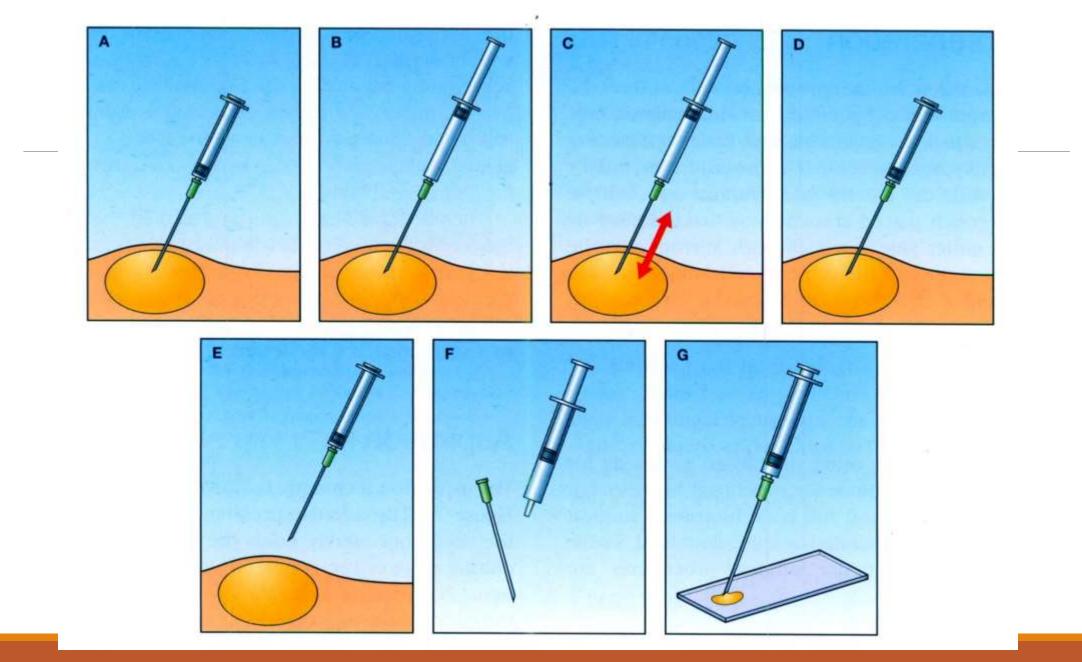


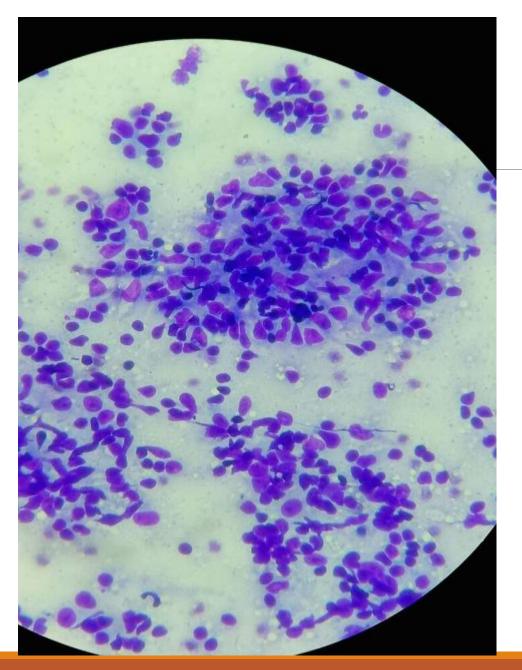


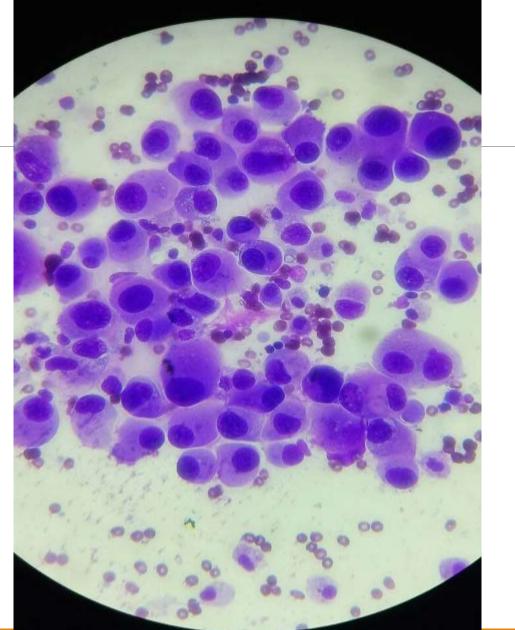












HYSTOCHEMISTRY/CYTOCHEMISTRY

Additional diagnostic tools

Untuk identifikasi komposisi bahan kimia yang

dihasilkan sel

			migal / av ta ab amigal ataina in
Та		tumour diagnosis.	nical/cytochemical stains in
	SUBSTAN	NCE	STAIN
1.	Basemen collagen	t membrane/	Periodic acid-Schif (PAS) Reticulin Van Gieson Masson's trichrome
2.	Glycogen	ו	PAS with diastase loss
3.		teins, glycolipids, cins (epithelial	PAS with diastase persist ence
4.	Acid muc (mesench	cin hymal origin)	Alcian blue
5.	Mucin (in	general)	Combined Alcian blue-PAS
6.	Argyroph argentaf	nilic⁄ n granules	Silver stains
7.	Cross stri	ations	PTAH stain
8.	Enzymes		Myeloperoxidase Acid phosphatase Alkaline phosphatase
9.	Nucleola regions(l	r organiser NORs)	Colloidal silver stain

	Tab	le 7.13 Common pane tumours of unc	l of immunohist ochemical stains for certain origin.
		TUMOUR	IMMUNOSTAIN
IMMUNOHISTOCHEMISTRY FUNGSI: • TUMOR OF UNCERTAIN HISTOGENESIS (DIAGNOSIS)	1.	Epithelial tumours (Carcinomas)	 i) Pankeratin (fractions: high and low molecular weight keratins, HMW-K, LMW-K) ii) Epithelial membrane antigen (EMA) iii) Carcinoembryonic antigen (CEA) iv) Neuron-specif c enolase (NSE)
 KATEGORISASI UNDIFF MALIGNANT TUMOR ASAL TUMOR METASTASIS PROGNOSTIC MARKER PREDIKSI RESPON TERAPI INFEKSI 	2.	Mesenchymal tumours (Sarcomas)	 i) Vimentin (general mesenchymal) ii) Desmin (for general myogenic) iii) Muscle specif c actin (for general myogenic) iv) Myoglobin (for skeletal myogenic) v) α-1-anti-chymotrypsin (for malignant f brous histiocytoma) vi) Factor VIII (for vascular tumours) vii) CD34 (endothelial marker)
	3.	Special groups	
		a) Melanoma	i) HMB-45 (most specif c) ii) Vimentin iii) S-100
		b) Lymphoma	 i) Leucocyte common antigen (LCA/ CD45) ii) Pan-B (Immunoglobulins, CD20) iii) Pan-T (CD3) iv) CD15, CD30 (RS cell marker for Hodgkin's)
		c) Neural and neuro- endocrine tumours	i) Neurof laments (NF) ii) NSE iii) GFAP (for glial tumours) iv) Chromogranin (for
			neuroendocrine)

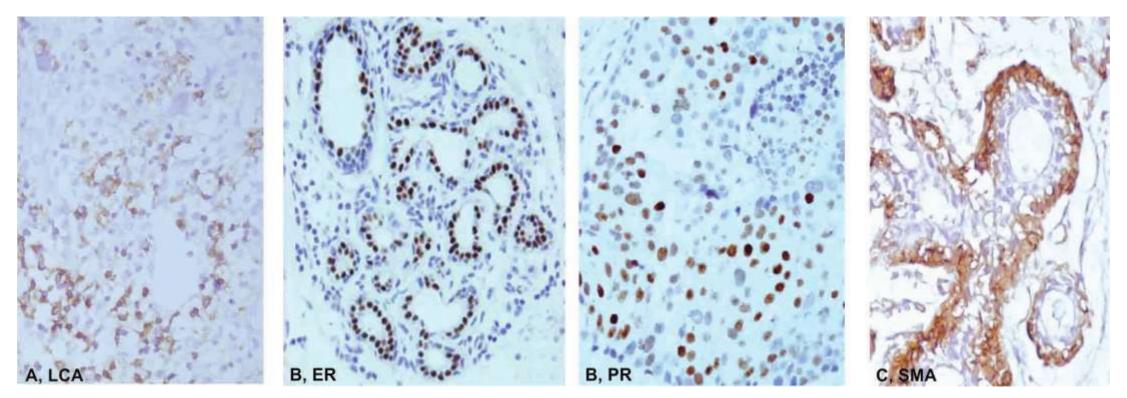


Figure 7.32 Examples of IHC staining at different sites in the tumour cells. A, Membranous staining for leucocyte common antigen (LCA) or CD45 in lymphomas. B, Cytoplasmic staining for smooth muscle actin (SMA) in myoepithelium on breast acinus. C, Nuclear staining for breast ER-PR receptor studies in breast cancer.

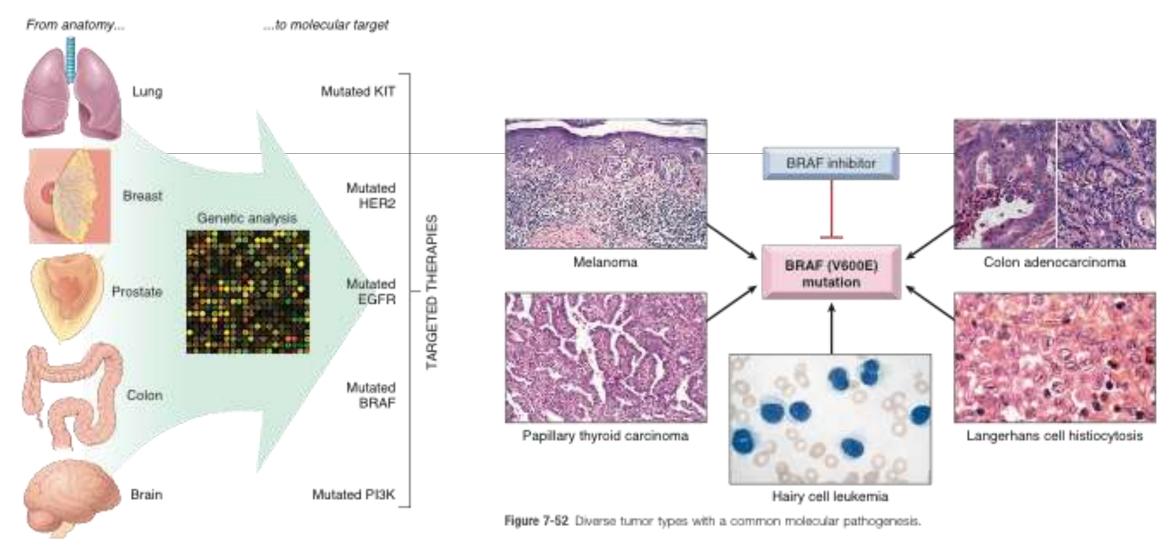


Figure 7-51 A paradigm shift: classification of cancer according to therapeutic targets rather than cell of origin and morphology. (Courtesy Dr. Levi Garraway, Dana Farber Cancer Institute.)

ELECTRON MISCROSCOPY

i) Cell junctions, their presence and type.

- ii) Cell surface, e.g. presence of microvilli.
- iii) Cell shape and cytoplasmic extensions.
- iv) Shape of the nucleus and features of nuclear membrane.
- v) Nucleoli, their size and density.
- vi) Cytoplasmic organelles—their number is generally reduced.
- vii) Dense bodies in the cytoplasm.

viii) Any other secretory product in the cytoplasm e.g. melanosomes in melanoma and membrane-bound granules in endocrine tumours.

TUMOR MARKER (BIOCHEMICAL ASSAY)

Tab	ble 7.14 Important tumour markers.	
	MARKER	CANCER
1.	ONCOFOETAL ANTIGENS	
	i. Alpha-foetoprotein (AFP) ii. Carcinoembryonic antigen (CEA)	Hepatocellular carcinoma, non-seminomatous germ cell tumours of testis Cancer of bowel, pancreas, breast
2.	Enzymes	
	<i>i. Prostate acid phosphatase (PAP) ii. Neuron-specif c enolase (NSE) iii. Lactic dehydrogenase (LDH)</i>	Prostatic carcinoma Neuroblastoma, oat cell carcinoma lung Lymphoma, Ewing's sarcoma
3.	HORMONES	
	i. Human chorionic gonadotropin (hOG) ii. Calcitonin iii. Catecholaminesand vanillyImandelic acid (VMA) iv. Ectopic hormone production	Trophoblastic tumours, non-seminomatous germ cell tumours of testis Medullary carcinoma thyroid Neuroblastoma, pheochromocytoma Paraneoplastic syndromes
4.	CANCER ASSOCIATED PROTEINS	
	 i. CA-125 ii. CA 15-3 iii. CA 19-9 iv. CD30 v. CD25 vi. Monoclonal immunoglobulins vii. Prostate specif c antigen (PSA) 	Ovary Breast Colon, pancreas, breast Hodgkin's disease, anaplastic large cell lymphoma (ALCL) Hairy cell leukaemia (HCL), adult T cell leukaemia lymphoma (ATLL) Multiple myeloma, other gammopathies Prostate carcinoma

OTHER MODERN AIDS

FLOW CYTOMETRY

IN SITU HYBRIDISATION (FISH/CISH)

CELL PROLIFERATION ANALYSIS (mitotic count, Ki-67, MIB)

IMAGE ANALYZER AND MORPHOMETRY

MOLECULAR DIAGNOSTIC TECHNIQUE

DNA MICROARRAY ANALYSIS TUMOR

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Formulir PA

Harus Berisi:

- No PA
- Identitas Pasien dan dokter pengirim
- Lokasi tumor (kp dengan gambar)
- Diagnosis Klinis
- Keterangan Kinis lainnya



HASIL PEMERIKSAAN PA

MAKROSKOPIS

Diterima 1 potong jaringan berat 260 gram, ukuran 15x12x5 cm, dilapisi kulit. Pada irisan tampak massa putih abu-abu ukuran 2,5x2x2 cm, putih abu-abu, padat kenyal. Jarak massa dengan dasar 1 cm. Jarak massa dengan kulit 1,5 cm. Pada eksplorasi tidak didapatkan pembesaran nodul KGB.

MIKROSKOPIS

Sediaan menunjukkan potongan jaringan mamm dengan proliferasi epitel anaplasi, inti bulat oval, pleomorfik berat, hiperkromatik, tersusun solid (tubular formation < 10%), tumbuh invasive ke dalam stroma. Mitosis 18/10 HpF. Papilla mamma dalam batas normal. Didapatkan angioinvasi: Jarak tumor dengan dasar (fascia) berhimpit Jarak tumor dengan kulit 10 mm.

KESIMPULAN

Mamma Dekstra, Operasi

INVASIVE CARCINOMA (IDC), NO SPECIAL TYPE, WHO GRADE III DIAMETER TUMOR 2.5 CM JARAK TUMOR DENGAN DASAR (FASCIA) BERHIMPIT (< 1 MM) JARAK TUMOR DENGAN KULIT 10 MM DIDAPATKAN ANGIONVASI PADA EKSPLORASI KGB TIDAK DITEMUKAN PEMBESARAN NODUL pT₃N₀M_{*}



Ir. Dian Y. Lestari, Sp.PA

FIKSASI

Tindakan merendam bahan pemeriksaan yang berasal dari jaringan tubuh kedalam cairan fiksasi

Tujuan Fiksasi:

- Mencegah terjadinya proses autolisis
- Mencegah proses pembusukan
- Memadatkan dan mengeraskan agar mudah dipotong
- Memadatkan cairan koloid
- Mencegah kerusakan struktur jaringan

JENIS FIKSASI

Untuk cairan \rightarrow urine, cairan pleura, bronchial washing

• alkohol 50% dengan volume perbandingan 1:1

Untuk sputum tampung

• Alkohol 70%

Bahan pap smear

• Alkohol 96%

Bahan Histopatologi

• Formaline 10% (buffer formalin), volume 5-10x ukuran jaringan

TRIMS

